

involving the use of indwelling catheters or transvenous pacing leads and cardiopulmonary bypass operations⁸.

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References

1. Minniti S, Vincentini S, Procacci C. Congenital anomalies of the venae cavae: embryological origin, imaging features and report of three new variants. *Eur Radiol* 2002; **12**: 2040–2055.
2. Nsah EN, Moore GW, Hutchins GM. Pathogenesis of persistent left superior vena cava with a coronary sinus connection. *Pediatr Pathol* 1991; **11**: 261–269.
3. Pasquini L, Belmar C, Seale A, Gardiner HM. Prenatal diagnosis of absent right and persistent left superior vena cava. *Prenat Diagn* 2006; **26**: 700–702.
4. Römer S, Opgen-Rhein B, Chaoui R, Scheer I, Czernik C, Obladen M. Bilateral agenesis of the superior vena cava associated with congenital hydrothorax. *Ultrasound Obstet Gynecol* 2006; **28**: 842–844.
5. Huhta JC, Smallhorn JF, Macartney FJ, Anderson RH, de Leval M. Cross-sectional echocardiographic diagnosis of systemic venous return. *Br Heart J* 1982; **48**: 388–403.
6. Guarnieri GF, Romano F, Clerico L, Balducci G. Absent right and persistent left superior vena cava: fetal and neonatal echocardiographic diagnosis. *Pediatr Cardiol* 2006; **27**: 646–648.
7. Chaoui R, Heling KS, Kalache KD. Caliber of the coronary sinus in fetuses with cardiac defects with and without left persistent superior vena cava and in growth-restricted fetuses with heart-sparing effect. *Prenat Diagn* 2003; **23**: 552–557.
8. Lenox CC, Zuberbuhler JR, Park SC, Neches WH, Mathews RA, Fricker FJ, Bahnsen HT, Siewers RD. Absent right superior vena cava with persistent left superior vena cava: implications and management. *Am J Cardiol* 1980; **45**: 117–122.

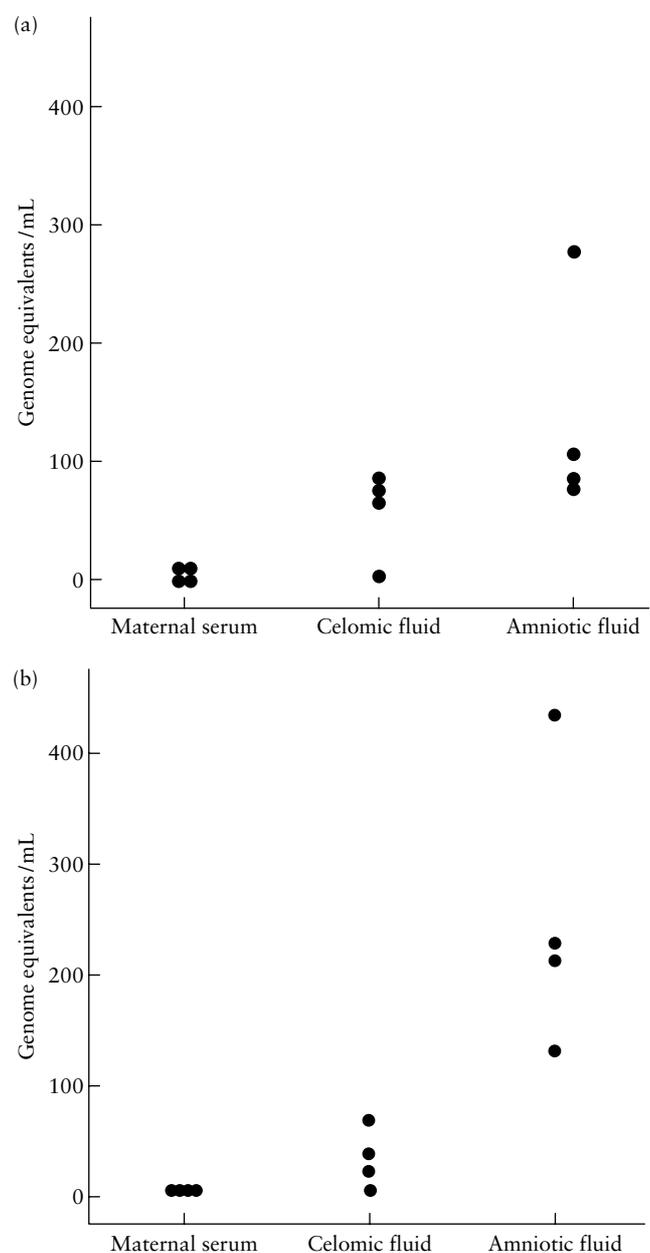
Cell-free fetal DNA in celomic fluid

Pregnancy is associated with high levels of cell-free DNA (cfDNA) in the maternal circulation, and around 5% of the DNA is fetal in origin¹. The exact source of cfDNA is uncertain, with some evidence supporting a placental^{2,3} and other a fetal⁴ origin. We further investigated the possible source of fetal cfDNA in maternal blood by comparing its concentration in maternal serum, amniotic fluid and celomic fluid.

Celocentesis was performed immediately before elective termination of pregnancy for psychosocial reasons in 11 singleton pregnancies with live fetuses at 7–9 weeks of gestation, as previously described⁵. Maternal blood (2 mL) was also obtained before the procedure. All participants were examined at the Department of Obstetrics and Gynaecology, University Hospital of Ioannina, Greece. The study was approved by the Hospital Ethics Committee, and written consent was

obtained. Male cfDNA was quantified by the SRY and the DYS14 gene sequences as previously described^{6,7}. The 'housekeeping' β -globin sequence was used to detect and quantify total free DNA¹, levels of which were expressed in genome equivalents (GE) per mL.

The analysis was restricted to the four male fetuses for which amniotic fluid was also available. Fetal cfDNA was demonstrated in all four samples of celomic fluid (SRY: median 71.5, range 3.0–75.1 GE/mL; DYS14: median 31.0, range 6.1–69.3 GE/mL) and amniotic fluid (SRY: median 96.6, range 79.0–279.1 GE/mL; DYS14: median 221.2, range 131.9–434.3 GE/mL), in all four maternal serum samples with the DYS14 marker (DYS14: median 5.8, range 5.5–7.4 GE/mL) and in two of the four sera using the SRY marker (Figure 1). No fetal cfDNA was demonstrated using the Y-chromosome specific markers



in either the celomic fluid or the maternal serum in four female fetuses that served as controls. Total cfDNA (β -globin DNA) was consistently found in all samples.

The concentration gradient of fetal cfDNA between the amniotic fluid (highest), celomic fluid (lower) and maternal serum (lowest) would seemingly indicate that the fetus itself is the source of fetal cfDNA in the maternal serum. However, it is very unlikely that fetal cfDNA from the early amniotic fluid could reach the maternal circulation because exchange at the level of the yolk sac is mainly unidirectional *towards* the fetus and the amniotic membrane is practically impermeable to large molecules⁸. Given that the estimated size of fetal cfDNA in maternal circulation is 100–300 base pairs⁹, which is equivalent to about 30–90 kDa, and the transfer through the amniotic membrane is negligible for molecules of similar size (e.g. human albumin), it appears that the source of cfDNA in the amniotic fluid is the fetus and the source in the maternal serum is the placenta. The alternative mechanism of direct transfer of fetal cfDNA between the fetal and maternal circulations is also unlikely because the uteroplacental circulation is established only after 10 weeks' gestation¹⁰. In support of this, a recent study found that the concentration of cfDNA in maternal serum was similar in normal and anembryonic pregnancies, indicating that the trophoblast is the source of cfDNA in maternal blood, even in the absence of a fetus or fetoplacental circulation³.

The robustness of our results is restricted by the small sample size, which mostly arose from the exclusion of most of the samples of the initial pool (female fetuses, $n = 4$; male fetuses without amniotic fluid, $n = 3$).

In summary, the *SRY* and *DYS14* markers are detected in the fluid compartments of very early pregnancy. Their distribution in the maternal serum and fetal fluid compartments provides preliminary evidence that the most likely source of fetal cfDNA in the maternal circulation is the trophoblast. A larger sample would be required in order to draw definite conclusions.

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References

- Lo YM, Tein MS, Lau TK, Haines CJ, Leung TN, Poon PM, Wainscoat JS, Johnson PJ, Chang AM, Hjelm NM. Quantitative analysis of fetal DNA in maternal plasma and serum: implications for noninvasive prenatal diagnosis. *Am J Hum Genet* 1998; **62**: 768–775.
- Guibert J, Benachi A, Grebille AG, Ernault P, Zorn JR, Costa JM. Kinetics of *SRY* gene appearance in maternal serum: detection by real time PCR in early pregnancy after assisted reproductive technique. *Hum Reprod* 2003; **18**: 1733–1736.
- Alberry M, Maddocks D, Jones M, Abdel Hadi M, Abdel-Fattah S, Avent N, Soothill PW. Free fetal DNA in maternal plasma in anembryonic pregnancies: confirmation that the origin is the trophoblast. *Prenat Diagn* 2007; **27**: 415–418.
- Ohashi Y, Miharu N, Honda H, Samura O, Ohama K. Correlation of fetal DNA and human chorionic gonadotropin concentrations in second-trimester maternal serum. *Clin Chem* 2002; **48**: 386–388.
- Makrydimas G, Georgiou I, Kranas V, Zikopoulos K, Lolis D. Prenatal diagnosis of beta-thalassaemia by coelocentesis. *Mol Hum Reprod* 1997; **3**: 729–731.
- Zimmermann B, El-Sheikhah A, Nicolaidis K, Holzgreve W, Hahn S. Optimized real-time quantitative PCR measurement of male fetal DNA in maternal plasma. *Clin Chem* 2005; **51**: 1598–1604.
- Gerovassili A, Garner C, Nicolaidis KH, Thein SL, Rees DC. Free fetal DNA in maternal circulation: a potential prognostic marker for chromosomal abnormalities? *Prenat Diagn* 2007; **27**: 104–110.
- Jauniaux E, Gulbis B. Fluid compartments of the embryonic environment. *Hum Reprod Update* 2000; **6**: 268–278.
- Chan KC, Zhang J, Hui AB, Wong N, Lau TK, Leung TN, Lo KW, Huang DW, Lo YM. Size distributions of maternal and fetal DNA in maternal plasma. *Clin Chem* 2004; **50**: 88–92.
- Burton GJ, Jauniaux E. Placental oxidative stress: from miscarriage to preeclampsia. *J Soc Gynecol Investig* 2004; **11**: 342–352.

Sonographic findings of uterine sacculation during pregnancy

Uterine sacculation is a rare complication of pregnancy that occurs in about 1 in 3000 pregnancies¹. It is defined as a sac-like structure that develops from an abnormal rotation of the uterine fundus^{2–4}. Without diagnosis and treatment, this can cause spontaneous miscarriage, intrauterine fetal death, uterine rupture, preterm delivery, placenta accreta, retained placenta and postpartum hemorrhage⁵. However, this condition can easily be misdiagnosed as uterine septum on ultrasound examination. We describe a case of uterine sacculation in a pregnancy which was further complicated by the presence of a large uterine myoma.

A 28-year-old primigravida presented with vaginal leakage and urinary retention at 17 weeks' gestation. Her medical history was unremarkable except for a known uterine myoma. On physical examination, we were unable to visualize the cervix with a speculum but bulging membranes were seen. A nitrazine test showed a positive result, indicating rupture of membranes. On ultrasound examination the fetus was found to be in breech-oblique presentation with a normal heartbeat and of normal size. However, precise evaluation of the fetus was limited owing to severe oligohydramnios. Ultrasonography showed probable uterine folding and a huge mass measuring around 10 × 8 cm in the upper portion of the uterus (Figure 1a). The anterior wall of the uterus was thin and stretched, but the posterior